3. 37

1. 2-8 20

2.

3. 0 S0 5 5

4.

5.

| | | | |
|---|---|---|---|
| | 96 | 48 | |
| | 12 ×8 | 12 ×4 | |
| | 0.3mL*6 | 0.3mL*6 | |
| | 6mL | 3mL | |
| -HRP | 10mL | 5mL | |
| 20× | 25mL | 15mL | |
| A | 6mL | 3mL | |

| | | |
|---|---|---|
| B | 6 mL | 3 mL |
| | 6 mL | 3 mL |
| | 2 | 2 |
| | 1 | 1 |
| | 1 | 1 |

4. HRP 100μL 37
60min

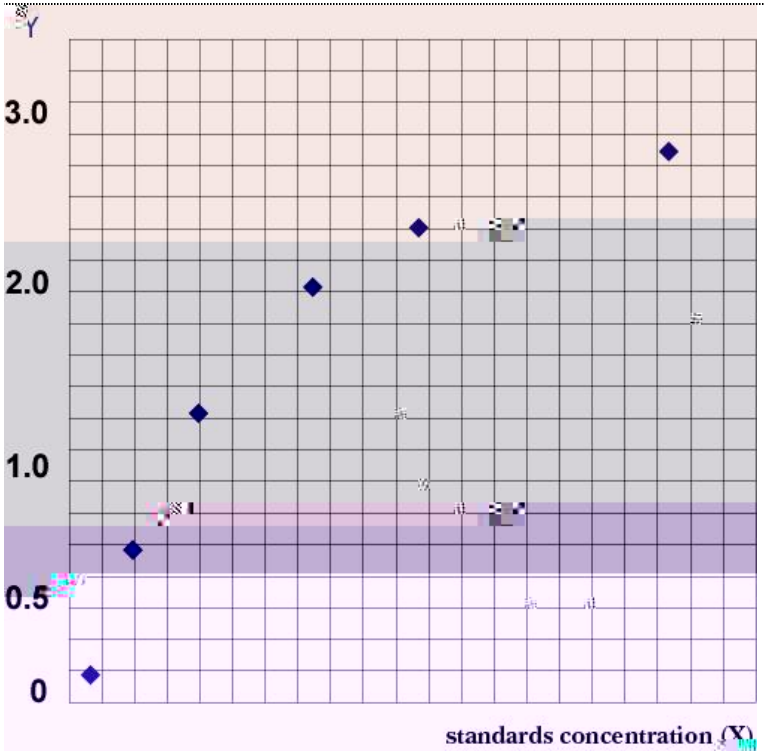
5. 1min 5

6. A B 50μL 37 15min

7. 50μL 15min 450nm OD

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Excel OD




1. R 0.9900

2. 10.0 U/L

3.

4. 15%

5. 2-8

6. 6

---

1.

2.

**FOR RESEARCH USE ONLY.**
**NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

# Plant Fructokinase (FK) ELISA Kit instruction

## Intended use

This FK ELISA kit is intended Laboratory for Research

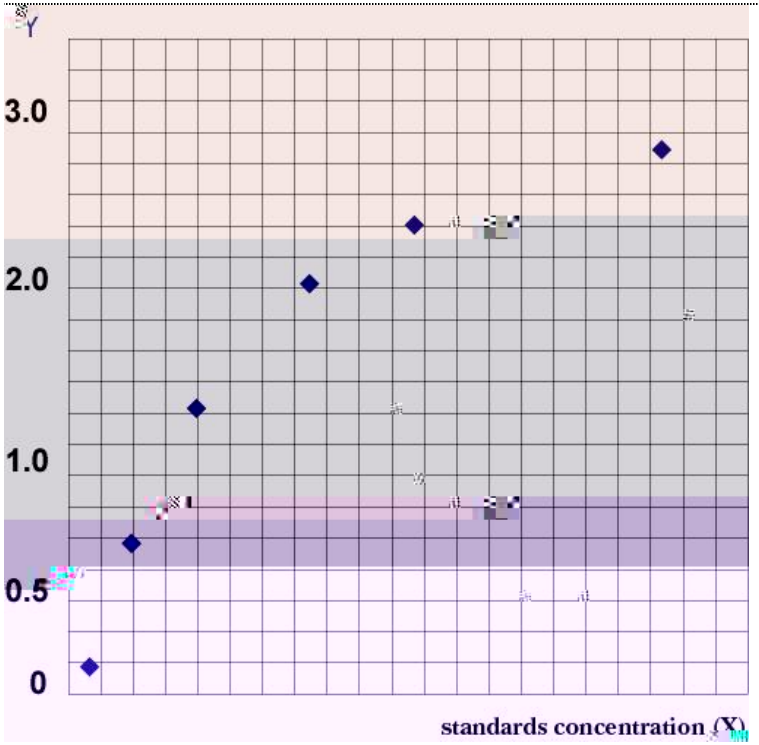
## Materials supplied

| Name | 96 determinations | 48 determinations |
|---|---|---|
| Microelisa stripplate | 12*8strips | 12*4strips |
| Standard | 0.3ml*6tubes | 0.3ml*6tubes |
| Sample Diluent | 6.0ml | 3.0ml |
| HRP-Conjugate reagent | 10.0ml | 5.0ml |
| 20X Wash solution | 25ml | 15ml |
| Chromogen Solution A | 6.0ml | 3.0ml |
| Chromogen Solution B | 6.0ml | 3.0ml |
| Stop Solution | 6.0ml | 3.0ml |

2. Add standard: Set Standard wells, testing sample wells. Add standard 50εl to standard well.
3. Add Sample: Add testing sample 10εl then add Sample Diluent 40εl to testing sample well; Blank well doesnôt add anyting.
4. Add 100εl of HRP-conjugate reagent to each well, cover with an adhesive strip and incubate for 60 minutes at 37ÁC.
5. Aspirate each well and wash, repeating the process four times for a total of five washes. Wash by filling each well with Wash Solution (400εl) using a squirt bottle, manifold dispenser or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add chromogen solution A 50εl and chromogen solution B 50εl to each well. Gently mix and incubate for 15 minutes at 37ÁC. **Protect from light.**
7. Add 50εl Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green or the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
8. Read the Optical Density (O.D.) at 450 nm using a microtiter plate reader within 15 minutes.

## Calculation of results

1. This standard curve is used to determine the amount in an unknown sample. The standard curve is generated by plotting the average O.D. (450 nm) obtained for each of the six standard concentrations on the vertical (Y) axis versus the corresponding concentration on the horizontal (X) axis.
2. First, calculate the mean O.D. value for each standard and sample. All O.D. values, are subtracted by the mean value of the zero standard before result interpretation. Construct the standard curve using graph paper or statistical software.
3. To determine the amount in each sample, first locate the O.D. value on the Y-axis and extend a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the corresponding concentration.
4. Any variation in operator, pipetting and washing technique, incubation time or temperature, and kit age can cause variation in result. Each user should obtain their own standard curve.
5. The sensitivity by this assay is 10.0 U/L
6. Standard curve




Storage 2-8 .
validity six months.

**FOR RESEARCH USE ONLY; NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS! PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING!**